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INTRODUCTION

The macrophage cytotoxic response is dependent on oxygen (O_2) as a substrate for oxidant production. Indeed, O₂ is utilized by (the inducible) nitric oxide synthase (iNOS) and NADPH oxidase-2 (NOX-2), to produce nitric oxide (•NO) and superoxide $(O_2^{\bullet-})$, respectively (Figure 1). Furthermore, the combination reaction between these species leads to peroxynitrite (ONOO⁻), a strong oxidant that can cause biological damage through oxidation and nitration. Since oxygen concentration varies between tissue and culture conditions, it has been of great interest to study these processes *in cellula*, considering the physiological environment. Previous reports from our group showed that oxidant production depends on oxygen partial pressure (pO_2) when evaluated during short-time exposures (Figure 2). Herein, we show the effects of long-term exposure to a range of physiological pO₂ on the activity and expression of iNOS.

Peroxynitrite formation, assessed through the boronate-based probe, coumarin-boronic ester (CBE), also shows a similar tendency with pO_2 in short-term exposures (scheme 1). Nevertheless, CBE is limited to only certain methodologies because of its spectroscopic properties. Moreover, other probes such as fluorescein-boronate (FI-B) are sensitive to drastic variations in intracellular pH and pO₂. Therefore, we are developing a novel fluorescent boronate-based probe (Red-B) derived from xanthene, to achieve an accurate cellular detection of peroxynitrite in different cellular conditions.



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Oxygen levels modulation of the macrophage oxidative response: nitric oxide and peroxynitrite production

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in 3 steps. B) Spectroscopic characterization of Red-Ox, showing its emission and excitation spectra and their differences with its protonated form (Red-Ox)⁺. **C)** Fluorescence dependence on pH for Fluorescein (FI) and Red-Ox.